

The characteristics and biological relevance of inorganic amorphous calcium carbonate (ACC) precipitated from seawater

David Evans^{1,a*}, Paul B. Webb², Kirsty Penkman³, Roland Kröger⁴, and Nicola Allison¹

¹ School of Earth and Environmental Sciences, University of St Andrews, St Andrews, UK

² School of Chemistry, University of St Andrews, St Andrews, UK

³ BioArCh, Department of Chemistry, University of York, York, UK

⁴ Department of Physics, University of York, York, UK

^a Now at: Institute for Geosciences, Goethe University Frankfurt, 60438 Frankfurt am Main, Germany

*evans@em.uni-frankfurt.de

Abstract

The importance of amorphous calcium carbonate (ACC) as a potential precursor phase in the biomineralization of marine calcifiers is increasingly being reported, particularly as the presence of ACC has been observed or inferred in several major groups. Here, we investigate the structure and conditions required to precipitate ACC from seawater-based solutions, with emphasis on the co-influence of the carbonate system (pH, dissolved inorganic carbon (DIC) concentration), seawater Mg/Ca ratio and the presence of amino acids. We find that Mg^{2+} and the presence of aspartic acid, glutamic acid, and glycine strongly inhibit ACC precipitation. Moreover, we were unable to precipitate ACC from seawater with a carbonate chemistry within the range of that thought to characterise the calcification site of certain marine calcifiers (i.e. $\text{DIC} < 6 \text{ mM}$, $\text{pH} < 9.3$), although substantial modification of the seawater Mg/Ca ratio ($\text{Mg}/\text{Ca}_{\text{sw}}$) allowed precipitation at a reduced DIC, with the implication that this could be an important component of utilising an ACC pathway. Finally, the degree to which $\text{Mg}/\text{Ca}_{\text{sw}}$ and the addition of amino acids influence the structure of ACC and the necessary seawater $[\text{CO}_3^{2-}]$ for precipitation is strongly pH dependent. At lower, more biologically relevant pH than that typical of much inorganic work, decreasing $\text{Mg}/\text{Ca}_{\text{sw}}$ can result in greater long-range order and less water of crystallisation, but facilitates precipitation at a considerably lower $[\text{CO}_3^{2-}]$ than at higher pH.

1. Introduction

Amorphous calcium carbonate (ACC) has been shown to play an integral role in the biomineralisation process in a wide range of organisms,^{1–4} including having been identified or inferred in a wide range of marine organisms including sponges,⁵ sea urchin larvae,⁶ ascidians,⁷ molluscs,⁸ coccolithophores,⁹ corals,¹⁰ and foraminifera.^{11–13} Improving our knowledge of the role of ACC in biomineralization is important because the shells of such organisms represent one of the largest long-term sinks of carbon on Earth,¹⁴ yet the effect of climate change and ocean acidification on the calcification process is not well-constrained,¹⁵ in part because some key aspects are poorly understood.

Inorganic ACC precipitation work has demonstrated that Mg^{2+} and organic molecules such as phosphoamino acids play a stabilising role,^{5,16,17} retarding the transformation of ACC into a crystalline carbonate. In addition, ACC may be stabilised in confinement,¹⁸ and the crystalline CaCO_3 polymorph that results from ACC depends on solution chemistry.^{19,20} Especially relevant to this study is the identification of synthesis-dependent structural variations in inorganic ACC,²¹ which has also been observed in biogenic samples. For example, Extended X-Ray Absorption Fine Structure (EXAFS) spectra of ACCs of different origin²² demonstrates not just variable Ca-O bond distances but also variable coordination numbers around Ca.

These studies yield insights into the diversity of ACC and controls on nucleation, growth and transformation, yet most inorganic precipitation work has utilised fluid chemistries which are very different from those inferred to be present at the site of ACC formation in marine calcifiers. For example, tropical zooxanthellate corals and hyaline foraminifera calcify from a seawater-like solution,^{11,23–26} albeit modified with respect to pH and DIC^{23,26–28}, whilst an ongoing debate surrounds the role of pumping protons, Ca^{2+} , and Mg^{2+} into and out of the calcifying space.^{13,29–31} In contrast, inorganic ACC is often experimentally precipitated through the decomposition of $(\text{NH}_4)_2\text{CO}_3$ in CaCl_2 solutions (Ref. 32 and references therein), or by mixing relatively concentrated equimolar CaCl_2 and Na_2CO_3 solutions.^{20,33,34} Both techniques result in solution carbonate chemistries that are highly variable throughout the course of the experiment, and far more saturated with respect to carbonate minerals compared to the calcifying space of marine organisms. For example, the calcifying space of corals is characterised by a DIC and pH of 2–6 mM and <9.3, respectively,^{26,28,35,36} a DIC concentration around two orders of magnitude lower than the initial solutions of e.g. Ref. 20. Furthermore, much previous ACC work was conducted in ionically simple solutions, raising the possibility that they might differ structurally and/or compositionally from biogenic ACCs. We stress that this is not intended as criticism of the large body of important work that exists, but rather highlights the need to build on these studies through the precipitation and study of ACC under conditions closer to those found in marine calcifiers.

We present data from a series of ACC precipitation experiments conducted in seawater-based solutions. Solution pH, DIC, $[\text{Mg}^{2+}]$, and $[\text{Ca}^{2+}]$ were varied to identify how these parameters affect the fluid saturation state required for ACC precipitation from seawater. We also studied the effect of aspartic and glutamic acid because the acidic amino acids are known to form ligands with Mg^{2+} and Ca^{2+} in solution,³⁷ and therefore may inhibit precipitation, as well as glycine for comparison. X-ray diffraction (XRD), Raman spectroscopy, attenuated total reflectance Fourier transform infrared spectroscopy (ATR-FTIR), and thermogravimetric analysis with coupled mass spectrometry (TGA-MS) were used to determine possible structural variations in the ACCs precipitated under different conditions.

2. Experimental

Sixty-six precipitation experiments were conducted spanning seawater Mg/Ca ratios of 0-5 mol mol⁻¹ (achieved by varying both Mg²⁺ and Ca²⁺), DIC concentrations of 3-25 mM, pH 8.9-10.3, as well as a range of L-aspartic acid, L-glutamic acid, and glycine (L-Asp, L-Glu, Gly) concentrations, and titration rates (see Tab. 1 and the Supplementary Information). We identified the solution saturation state required to precipitate ACC under these conditions, hereafter abbreviated to ACC_{SI} (Saturation Index):

$$\text{ACC}_{\text{SI}} = [\text{Ca}^{2+}] \cdot [\text{CO}_3^{2-}] \quad (\text{Eq. 1})$$

Where [Ca²⁺] and [CO₃²⁻] refer to the concentrations at the onset of ACC precipitation. ACC_{SI} is directly proportional to the CaCO₃ saturation state of the solution (Ω), where for example $\Omega_{\text{calcite}} = [\text{Ca}^{2+}] \cdot [\text{CO}_3^{2-}] / K_{\text{sp}}(\text{calcite})$. We use a simpler formulation of the solution saturation state because ACC K_{sp} is not well-known, and likely varies depending on the structure and composition of the ACC precipitated.³⁸ A higher ACC_{SI} directly implies a higher [Ca²⁺] and/or [CO₃²⁻] required to precipitate ACC from seawater.

2.1 ACC precipitation

All precipitation experiments were conducted in artificial seawater (ASW) made using the protocol given in Ref. 39, modified to include the most widely studied trace elements in biogenic carbonates at a concentration approximately equivalent to open-ocean seawater (see the Supplementary Information). The majority of experiments were conducted in seawater with the natural Mg/Ca ratio of 5.2 mol mol⁻¹ (53.0/10.3 mM/mM) whilst a subset of 20 investigated the effect of independently varying both [Mg²⁺] and/or [Ca²⁺]. The salinity of most experiments was 35 PSU, except where the [Mg²⁺] or [Ca²⁺] was modified. Seawater with a [Mg²⁺] lower than natural was produced by adding lower amounts of MgCl₂, such that in these experiments, salinity was proportional to [Mg²⁺] (33 PSU in MgCl₂-free ASW). Seawater with a [Ca²⁺] higher than natural (>10.3 mM) was produced by spiking with CaCl₂, such that salinity scaled linearly from 35 to 37 PSU in those with the highest [Ca²⁺] (50 mM).

To precipitate ACC, the carbonate chemistry of the seawater-based solutions was adjusted, by trial and error, to a state just below that required for ACC precipitation, i.e. ACC precipitation never occurred spontaneously but was initiated by subsequent dosing of small volumes of CaCl₂ and Na₂CO₃. The seawater pH and [Ca²⁺] were monitored using pH and Ca ion selective electrodes (ISE), calibrated using NIST-traceable buffer solutions and artificial seawater solutions respectively, the latter with a range of seawaters with [Ca²⁺] spanning 0-16 mM (determined by ICP-OES). Prior to ACC precipitation, the seawater carbonate chemistry was adjusted to the desired value by adding 1 M Na₂CO₃ (to increase DIC) and 0.1 M NaOH or HCl to adjust pH (Fig. 1A). The initial seawater DIC was calculated as the measured DIC of the stock seawater plus the amount of Na₂CO₃ added. Once the

desired solution chemistry was obtained, the seawater was stirred slowly for 60 s to allow the carbonate system to equilibrate. The Ca ISE was monitored to confirm that no precipitation occurred while the solution chemistry was being manipulated (Fig. 1B). ACC precipitation was then initiated by slow, controlled, simultaneous titration of 0.45 M CaCl_2 and Na_2CO_3 solutions into the seawater. All precipitations were conducted in acid-cleaned (1 M HCl) High-Density Polyethylene (HDPE) beakers at $22.5 \pm 1.5^\circ\text{C}$ using a Metrohm 902 Titrando titrator. The Ca^{2+} ISE has a minor sensitivity to pH (a 1 unit pH change results in an approximate seawater $[\text{Ca}^{2+}]$ bias of 15%) which was corrected for using the pH electrode.

The solution chemistry ($[\text{Ca}^{2+}]$, pH and [DIC]) conditions at the onset of ACC precipitation were identified by comparing the observed evolution of $[\text{Ca}^{2+}]$ using the calibrated Ca ISE to that predicted if all titrated CaCl_2 remained in solution (Fig. 1B). The seawater $[\text{Ca}^{2+}]$ at ACC_{SI} (the conditions at which precipitation began) is defined here as the point at which more than 20% of the titrated CaCl_2 was utilised in precipitation. Whilst this method slightly overestimates the solution saturation state with respect to carbonate precipitates (compare arrows 2 and 3 in Fig. 1C), this compromise was ultimately preferred over the uncertainty associated with precisely defining the point at which the Ca^{2+} titration curve deviated from expected, as the Ca ISE has a precision of $\sim 3\%$ in high-ionic strength solutions such as seawater. The seawater $[\text{CO}_3^{2-}]$ required to calculate ACC_{SI} was calculated from DIC and pH using *co2sys*,⁴⁰ based on the titration of exactly equal volumes of Na_2CO_3 and CaCl_2 up until the point that precipitation began, i.e. $\Delta\text{DIC} = \Delta\text{Ca}$. In these relatively short experiments (precipitation typically took place over <5 minutes, total experiment duration including seawater carbonate chemistry adjustment was <20 minutes), the seawater carbonate chemistry was assumed to be unaffected by CO_2 diffusion from the atmosphere.

The majority of experiments were conducted in 100-200 ml seawater, with a subset (20 precipitations) conducted in 250 ml. In these larger volume experiments seawater samples for DIC and elemental chemistry were taken, albeit only after sufficient precipitation had taken place for characterisation (30-60 s after ACC_{SI} , the point at which precipitation began), see the Supplementary Information for further details. The seawater-precipitate mixtures were immediately vacuum separated using $0.2 \mu\text{m}$ nylon membrane filters, and the DIC of the filtrate was immediately determined. Samples for elemental analysis were placed into acid-cleaned (1 M HCl) polypropylene centrifuge tubes and immediately acidified to 5% HNO_3 . The precipitates were thoroughly rinsed with trace element grade ethanol and air-dried. We observe good agreement between the measured Mg/Ca and DIC estimates based on the technique outlined in the preceding paragraph (typically better than 7% in both cases, see the Supplementary Information)

Based on the technique described above, we calculate the solution carbonate chemistry (pH, DIC, $[\text{CO}_3^{2-}]$), $[\text{Ca}^{2+}]$ and Mg/Ca ratio at the onset of precipitation, use these data to compute ACC_{SI} , and subsequently examine how this is impacted by the experimental variables described above (summarised in Tab. 1). The seawater Mg/Ca ratio at the onset of precipitation was calculated as $[\text{Mg}^{2+}_{\text{initial}}]/([\text{Ca}^{2+}_{\text{initial}}] + \text{Ca}^{2+}_{\text{titrated}})$, i.e. assuming the titrant solutions did not contain a significant Mg contaminant, and that only a minor amount of Mg^{2+} was removed through ACC precipitation. We estimate that precipitation of 20 mg ACC (typical of the 250 ml experiments) with a Mg concentration of 10 mole %⁴¹ reduced the solution $[\text{Mg}^{2+}]$ by 0.04 mM, or 0.4%, which we consider to be unimportant in the context of the large range investigated here (0-53 mM). Similarly, the relatively small titration volumes necessary to precipitate ACC using this method (~1-4 ml in 100-250 ml seawater) meant that there was no substantial change in ionic strength over the course of the experiments.

2.2 Analytical chemistry

The uncertainty in the measurement of pH was calculated as 0.020 across the pH range 4-10, based on the maximum observed difference between fresh and one-week-old pH buffers (the buffers used to perform the calibration were replaced approximately once per week).

Seawater DIC was analysed using a LI-7000 CO_2 differential, non-dispersive, infrared gas analyser (Apollo SciTech; AS-C3). The instrument was calibrated at intervals using a seawater standard (Dickson batch 141, DIC $2033.3 \pm 0.3 \text{ mol kg}^{-1}$), and the accuracy and linearity of the analyser at the relatively high DIC concentrations of this study was confirmed by analysis of a range of Na_2CO_3 solutions spanning 1.5-16 mM. The precision of individual samples assessed through multiple (6-10) injections was routinely better than 0.2%. However, because the DIC analyser was not always calibrated before each set of experiments, and significant variations in the DIC calibration were observed, we report a DIC uncertainty of $\pm 7\%$ based on the maximum difference between calibrations performed on different days.

$\text{Mg}/\text{Ca}_{\text{sw}}$ was determined using the Varian Vista Pro ICP-OES (axial) at the Edinburgh Earth Observatory. The samples were diluted 1:50 with 3% HNO_3 to reduce the plasma loading associated with high-ionic strength solutions. Yttrium was used as the internal standard. Calibration was performed using a six-point calibration line (blank plus five standards), spanning the range of sample Mg and Ca concentrations. Accuracy and precision (2RSD) of measured $\text{Mg}/\text{Ca}_{\text{sw}}$ were 1.8% and 2.2% respectively based on repeat analyses of natural seawater.

2.3 Precipitate characterisation

Raman, ATR-FTIR, XRD, and TGA-MS analysis were performed on a subset of the precipitates covering the range of experimental conditions. Raman spectroscopy measurements were performed using a Horiba Jobin Yvon LabRam HR800 with a 50× long-working-distance objective, an excitation wavelength of 514 nm, and beam diameter of 1 μm . The system was calibrated prior to each set of measurements using an Ag standard. Measurements were performed at 1.8 cm^{-1} resolution with the reported spectra representing the average of 20×5s scans. Initial measurements were performed at a low laser intensity to ensure that local heating on the sample did not alter the ACC. The laser intensity was then systematically increased to maximise signal/noise whilst ensuring no thermally induced transformation took place (i.e. no change in the spectra when normalised to the most intense peak). Subsequent full width at half maximum peak height (FWHM) analysis is dependent on spectral resolution,^{42,43} such that the relatively wide peak widths we report for crystalline CaCO_3 should be viewed in the context of the spectral resolution used here. In contrast, the disordered nature of ACC means that the peaks of interest are weakly dependent on spectral resolution (>20 wavenumbers wide) and easily distinguishable from crystalline materials (~4 times wider). ATR-FTIR spectra were collected using a Bruker Platinum ATR infrared spectrometer fitted with a TGS detector; measurements were performed at 1.4 cm^{-1} resolution with 32 scans. A baseline measurement was performed before every sample. TGA-MS profiles were determined using a Netzsch STA 449 F1 Jupiter coupled with a QMS 403 Aëolos R quadrupole mass spectrometer. Around 10 mg of sample was heated at a rate of 10°C min^{-1} from 40°C to 800°C, in an Ar atmosphere with a flow rate of 350 ml min^{-1} . As well as TGA profiles, the ion current of m/z 18, 28 and 44 was monitored to enable the identification of the decomposition products. In order to calculate the formula water of crystallisation, the relationship between $n\text{-H}_2\text{O}$ and the integrated area of the m/z 18 ion peak was calibrated based on the thermal decomposition of $\text{CaC}_2\text{O}_4\cdot\text{H}_2\text{O}$ (see the Supplementary Information).

3. Results and discussion

The solution chemistry conditions at ACC_{SI} (the onset of precipitation) is shown in pH-DIC space in Fig. 2. The majority of experiments fall within ± 2 mM of the 10 mM $[\text{CO}_3^{2-}]$ line (dashed contours overlain on Fig. 2), equivalent to a $[\text{Ca}^{2+}]/[\text{CO}_3^{2-}]$ ratio of ~1-1.2, with the exception of those in which the Mg/Ca ratio was modified. Given that the experimental seawaters did not have initial equimolar concentrations of $[\text{Ca}^{2+}]$ and $[\text{CO}_3^{2-}]$, this is not an artefact of the experimental design. Therefore, we find that in seawater with a Mg/Ca ratio close to natural (5 mol mol^{-1}), ACC precipitation occurs when the DIC concentration reaches a value necessary to achieve approximately equimolar seawater $[\text{CO}_3^{2-}]$ and $[\text{Ca}^{2+}]$.

3.1 Vibrational spectroscopy and thermal decomposition

Raman, FTIR, XRD and TGA-MS analyses (Fig. 3,4,5,6), confirm that our experiments yielded ACC in all but two cases. Experiments conducted in seawater with an initial Mg/Ca ratio <1 mol mol^{-1}

resulted in rapid calcite precipitation without an observed ACC precursor (e.g. Fig. 3F,5), while those conducted at the highest pH (10.3 on the NBS scale) resulted in amorphous brucite-CaCO₃ mixtures (Fig. 4H).

In the variable pH/DIC experiments, there is no resolvable control of solution carbonate chemistry on any aspect of the Raman and FTIR spectra when precipitation takes place from normal seawater (Mg/Ca = 53/10.3 mM/mM) at pH ~10. Whilst DIC and pH were varied across a wide range, most ACCs were precipitated at a [CO₃²⁻] of 10±2 mM and a Mg/Ca_{sw} ratio of ~5 mol mol⁻¹ (Fig. 2). This may explain why there is little evidence for a carbonate chemistry control on ACC structure, as the seawater [Ca²⁺]/[CO₃²⁻] ratio was close to one in most cases. Similarly, in the variable amino acid concentration experiments, we find no resolvable structural effect on ACC as evidenced by the FTIR or Raman spectra (Fig. 3 and 4), in agreement with a previous study.⁴⁴ These molecules have a differential affinity to form ligands with Ca²⁺ and Mg²⁺ in solution, such that amino acid concentration affects the free Mg/Ca ratio.³⁷ Therefore, it is somewhat surprising that the concentration of Asp and Glu do not resolvably impact ACC structure, as both influence Mg incorporation into ACC, and we identify Mg/Ca_{sw} as a driver of ACC structure (Fig. 7).

The Raman spectra of ACC (Fig. 3) are dominated by a broad ν_1 peak (CO₃ symmetric stretch) at 1080 cm⁻¹ (the precise location of this peak varies as a function of mole % Mg⁴²), and a smaller broad ν_4 peak (CO₃ in-plane bend) at 710 cm⁻¹. In ACC precipitated from seawater with the natural [Mg²⁺] and [Ca²⁺], the lattice mode vibrations of the crystalline CaCO₃ polymorphs, if present, are obscured due to the presence of a broad band below 300 cm⁻¹. This feature occurs in both biogenic ACC² and inorganic hydrated crystalline and amorphous CaCO₃ (Ref. 45), but not in the crystalline anhydrous polymorphs (e.g. Fig. 3F), indicating that it relates to the presence of water. In contrast, ACC precipitated from seawater with Mg²⁺ reduced from 53 to 10 mM (Fig. 3E) is characterised by spectra with narrower ν_1 and ν_4 peaks, as well as distinguishable lattice mode vibrations at 155 cm⁻¹ and 285 cm⁻¹, indicative of the crystal structure of calcite. These precipitates either differ structurally from their counterparts formed at higher Mg/Ca_{sw} or represent physical mixtures of calcite and ACC. Interestingly, the Raman spectra of the precipitates presented here show many of the different features reported for both stable and transient biogenic ACCs.² Given that the differences in the Raman spectra between our experiments are principally driven by the seawater Mg/Ca ratio, this raises the possibility that variation in the structure of biogenic ACCs also arise from the Mg/Ca ratio at the site of calcification, further discussed below.

The full width at half maximum (FWHM) of the symmetric C-O stretch (ν_1) peak at ~1080 cm⁻¹ of all ACCs is positively correlated with Mg/Ca_{sw} and greatly exceed that of calcite and aragonite (22-34 cm⁻¹ compared to 5.5-10.8 cm⁻¹ respectively, Fig. 7). Note that this comparison should be viewed in the context of the spectral resolution of our Raman data (~1.8 cm⁻¹), such that the FWHM of the ACC

samples may differ even more from calcite and aragonite than is apparent. The Root Mean Square Error (RMSE) of the ν_1 FWHM based on the data in Fig. 7 is 1.1 cm^{-1} , such that these ACCs are significantly different from the crystalline samples (as previously described⁴), and the relationship between $\text{Mg}/\text{Ca}_{\text{sw}}$ and FWHM of the ACCs is highly significant ($R^2 = 0.92$; $p < 0.01$). The relatively wide ACC ν_1 peak occurs because materials lacking long-range order are typically characterised by broader peaks, in the case of ACC due to the disordered coordination environment around the Ca atoms compared to calcite.⁴⁶ In addition, both the relatively large ν_1 peak width in ACC and the increase in ν_1 FWHM with $\text{Mg}/\text{Ca}_{\text{sw}}$ arise because ACC typically contains >10 mole % Mg.⁴² Because Mg-O has a longer metal-O bond distance than Ca-O, (increasing) Mg incorporation results in greater bulk heterogeneity in metal-O vibrations, and a wider ν_1 peak. In this respect, our data are in good agreement with those of Ref. 42 who found the ν_1 FWHM is linearly dependent on the mole % Mg of ACC. Whilst we cannot directly compare our results to that study as we do not report precipitate Mg/Ca data, it has been shown that ACC Mg/Ca is proportional to $\text{Mg}/\text{Ca}_{\text{sw}}$,⁴¹ such that our data are consistent with this finding.

Both one and two peak fits of the ν_1 band of amorphous precipitates formed under different $\text{Mg}/\text{Ca}_{\text{sw}}$ are shown in Fig. 7A. The position of the peaks in the two-peak fit were fixed at 1080 cm^{-1} and 1090 cm^{-1} (CaCO_3 and MgCO_3 respectively⁴⁷), whilst the peak widths were fixed at 20 cm^{-1} , the intercept of our $\text{Mg}/\text{Ca}_{\text{sw}}$ -FWHM regression (Fig. 7A), inferred to represent Mg-free ACC. A single peak fit is likely more appropriate if the ν_1 band is symmetrical and therefore explicable by solid solution of amorphous $(\text{Ca,Mg})(\text{CO}_3)_2$, whereas a two peak fit may be more appropriate if the ν_1 band is asymmetrical, which would imply a heterogeneous material. The residuals of the one and two-peak fit are similar for ACCs precipitated from different $\text{Mg}/\text{Ca}_{\text{sw}}$ ratios achieved by varying Ca^{2+} at constant Mg^{2+} (Fig. 7B-C). Therefore, the ν_1 band of these ACCs is (i) not resolvably asymmetric (cf. Ref. 42), and (ii) well-described by two closely-spaced peaks representing amorphous CaCO_3 and MgCO_3 endmembers; a solid-solution model of Mg incorporation into ACC is compatible with these data. In contrast, ACC precipitated from seawater with a Mg^{2+} concentration reduced from 53 to 10 mM ($\text{Mg}/\text{Ca}_{\text{sw}} \sim 1$; Fig. 7D) is characterised by an asymmetrical ν_1 band, which cannot be modelled by any combination of the two-peak fit. Rather, the best-fit two-peak model arises through the combination of a relative wide peak (20 cm^{-1}) centred on 1080 cm^{-1} and a much narrower (8 cm^{-1}) peak centred on 1086 cm^{-1} (this fit was determined by fixing the position and FWHM of the wider peak for the same reasons as above and solving for all other parameters, i.e. treating the second component as a complete unknown). The narrow peak indicates the presence of a more ordered phase, whilst the position of this peak is consistent with that of low-Mg calcite.⁴⁷ Thus, precipitation at a $[\text{Mg}^{2+}]$ greatly lower than normal seawater may result in ACC-calcite mixtures despite the overall highly saturated conditions investigated (Tab. 1). The implication of this is that within the DIC and pH range investigated here (as close as possible to that thought to characterise certain marine

calcifiers) it appears that there is a limit to which the $[\text{Mg}^{2+}]$ can be reduced before the resulting precipitate is not entirely ACC.

The most prominent feature of the FTIR spectra is the split ν_3 band at ~ 1400 and 1474 cm^{-1} (antisymmetric CO_3 stretch; Fig. 4), indicating that there is more than one local symmetry around the CO_3 ion in ACC,² and therefore that ACCs do have a short-range order. The broad band derived from the hydrogen-bonded O-H stretch between $3000\text{--}3800\text{ cm}^{-1}$ indicates the presence of a significant amount of water of crystallisation, confirmed by TGA-MS (Fig. 6A). The thermal decomposition profiles of ACC precipitated from normal seawater ($\text{Mg}/\text{Ca} = 53/10.3\text{ mM}/\text{mM}$) show a mass loss associated with the $m/z\ 18$ ion beam of 17-19%, equivalent to a formula mass of $\text{CaCO}_3 \cdot 0.85 \pm 0.05\text{H}_2\text{O}$. As in the case of the Raman and FTIR spectra, we find no systematic relationship between water content and seawater carbonate chemistry or amino acid concentration.

ACC precipitated from seawater with a $[\text{Mg}^{2+}]$ reduced to 10 mM has a narrower ν_2 peak at 864 cm^{-1} and a greatly reduced O-H band between $3000\text{--}3800\text{ cm}^{-1}$ (Fig. 4F). This further indicates that the Mg^{2+} concentration or ratio of Mg/Ca imparts the greatest effect on the structure of ACC and water of crystallisation (and/or results in ACC-crystalline mixtures at low $[\text{Mg}^{2+}]$ as discussed above; Fig. 7D). Moreover, although varying pH and DIC result in no resolvable structural differences in ACC precipitated from normal seawater, we find that this is not the case as $\text{Mg}/\text{Ca}_{\text{sw}}$ is reduced. At pH 10.1 and a seawater Mg/Ca ratio of 1 mol mol^{-1} , achieved by raising $[\text{Ca}^{2+}]$ from 10 to 50 mM, the resulting ACC has an FTIR spectra equivalent to those of precipitates from seawater with the natural Mg/Ca ratio (compare panels A and D in Fig. 4). However, when ACC is precipitated from seawater with 50 mM Ca^{2+} but a pH slightly reduced to 9.7, the resulting spectra are characterised by a narrower ν_2 peak at 864 cm^{-1} (FWHM reduced from 15.5 to 7.2 cm^{-1} , see Fig. 4D,E), as well as a narrower Raman ν_1 peak. The asymmetry of the ν_3 band, lack of a resolvable ν_1 peak in the FTIR spectra, and the presence of a significant amount of water of crystallisation indicate a dominantly amorphous precipitate. However, the bulk material has a greater degree of short-range order compared to ACC precipitated from normal seawater. Previous work has shown that the presence of ‘additives’ like Mg^{2+} impact the short-range structure of ACC^{21,48}; our results demonstrate that the extent to which the seawater Mg/Ca ratio drives structural differences is dependent on the carbonate chemistry. Specifically, the seawater Mg/Ca ratio exerts a larger influence on the structure of these materials at lower (more biologically relevant) pH. Given that $\text{Mg}/\text{Ca}_{\text{sw}}$ is the only investigated method by which the solution $[\text{Ca}^{2+}]/[\text{CO}_3^{2-}]$ ratio at the onset of precipitation can be greatly reduced below unity (Fig. 2), it may be that the mechanistic basis for the $\text{Mg}/\text{Ca}_{\text{sw}}$ -dependent structural variation in ACC is rooted in precipitation from solutions with a relatively low $[\text{CO}_3^{2-}]$. Of relevance here is the previous use of nuclear magnetic resonance (NMR) spectroscopy to identify structural differences in the position of mobile water in biological ACC compared to synthetic ACC produced at high pH.⁴⁹ We

find that the seawater Mg/Ca ratio is the only parameter we varied that resulted in a substantial variation in the precipitate water of crystallisation (Fig. 4). Together, these observations again point to Mg/Ca_{sw} as a key factor that could result in structural differences in both the precipitates discussed here as well as biogenic ACCs.

3.2 Solution chemistry control on ACC precipitation from seawater

The impacts of the investigated variables on ACC_{SI} (Eq. 1) are summarised in Fig. 8 and 9. Because some factors unavoidably covaried in our experiments (especially solution [Ca²⁺] with other variables), the experiments were grouped into five broad sets on which to perform least-squares multiple linear regression analyses (Tab. 2). The aim of this exercise was to identify which factors (the carbonate system, solution [Mg²⁺] and/or [Ca²⁺], and amino acid concentration) exert the largest control on ACC_{SI} where these vary together, and to explore how much of the variance in the data can be explained by these variables alone. A linear model of the following form was chosen for simplicity:

$$\text{ACC}_{\text{SI}} = x_1 + x_2 \text{Mg/Ca}_{\text{sw}} + x_3 \text{pH} + x_4 \text{DIC} + x_5 [\text{AA}] \quad (\text{Eq. 2})$$

However, we note that there is no theoretical basis for this, and more complex models could result in better fits to the data if ACC_{SI} is nonlinearly sensitive to some factors. pH and DIC were used to account for the carbonate system because these were the variables that were measured, but ACC_{SI} may be mechanistically more appropriately related to (e.g.) pH and [CO₃²⁻] (albeit there is no significant difference in the model fits if [CO₃²⁻] rather than DIC is chosen).

All multiple linear regression models are highly significant (Tab. 2, $p < 0.01$), except for the glutamic acid experiment which contained far fewer precipitations. This exercise demonstrates that solution pH, DIC, [Mg²⁺], [Ca²⁺], and amino acid concentration can explain nearly all the variance in the data (model R² range between 0.79-0.98, RMSE 5.2-9.3). Grouping all data together results in a model in which all independent variables except for amino acid concentration are significant predictors of ACC_{SI}, with ACC_{SI} being most sensitive to pH ($m = \sim 50$ per unit), followed by Mg/Ca_{sw} ($m = \sim 30$ per mol/mol).

The relationship between DIC and ACC_{SI} is not significant when DIC alone is considered (Fig. 8A), yet the multiple linear regression models demonstrate that in the amino acid-free experiments DIC is a significant predictor of ACC_{SI} both at normal and low seawater Mg concentrations, and when the entire dataset is combined (Tab. 2). This relationship is significant irrespective of whether our best estimate of ACC_{SI} is used, or if the uncertainty in ACC_{SI} (mostly derived from the DIC calibration) is fully propagated through to the model by Monte Carlo simulation (1000 simulations per model, see the SI for details), because almost all of the uncertainty in ACC_{SI} is derived from the DIC uncertainty (the uncertainties are almost perfectly correlated). As a result, the uncertainty in absolute DIC has a

very minor effect on the multiple linear regression models, and we are therefore able to identify the carbonate system as a significant factor influencing ACC precipitation from seawater. Specifically, increasing DIC and/or pH increases ACC_{SI} . All else being equal this would imply a higher $[CO_3^{2-}]$ necessary for ACC precipitation, which indicates that some factor slightly inhibits precipitation under these overall more saturated conditions. Potentially relevant considerations are pH or concentration-dependent ion-pairing, or the size of hydration spheres.

Titration rate was not found to impart a systematic control on ACC_{SI} (Fig. 8D), and this factor was not investigated further. Importantly, however, this result does indicate that varying titration rate by more than an order of magnitude does not greatly affect the identification of ACC_{SI} using this experimental design.

High concentrations of the acidic amino acids substantially inhibit ACC precipitation (Fig. 8E,F), so it is surprising that the ACC_{SI} slope associated with this factor in the multiple linear regression models is relatively minor ($m = 0.1-0.6$ per mM; Tab. 2). The reason for this may be that the inhibitory effect of amino acids in seawater is principally overcome through the addition of Ca^{2+} in our experiments (Fig. 1C), so that ACC_{SI} is ultimately best correlated with the seawater Mg/Ca ratio. To test this, the linear regression models were run both with and without amino acid concentration as an independent variable (Tab. 2). In the amino-acid free models Mg/ Ca_{sw} becomes a significant predictor of ACC_{SI} , which is to be expected in the case of collinearity in the independent variables, adding support to the above hypothesis. That is, the non-significance of Mg/ Ca_{sw} as a predictor of ACC_{SI} in the amino acid experiments when both [AA] and Mg/ Ca_{sw} are used in the regression model results from the tight correlation between [AA] and Mg/ Ca_{sw} at the onset of precipitation, as the inhibitory effects of the amino acids is overcome through the additional titration of Ca^{2+} .

The role of Mg^{2+} in both inhibiting ACC precipitation and stabilising against transformation to a crystalline $CaCO_3$ is well-known.^{2,50,51} It is therefore unsurprising that we find it also plays a similarly important role in ACC precipitation from seawater. Reducing seawater $[Mg^{2+}]$ from 53 to 10 mM enables precipitation at a moderately lower ACC_{SI} , and therefore a lower DIC, given the initial seawater $[Ca^{2+}]$ was not varied in these experiments (Fig. 8B). By conducting a set of experiments at different pH and DIC with $[Mg^{2+}]$ reduced to 10 mM, we show that it is possible to precipitate ACC at a DIC below 10 mM (Fig. 8B,C), i.e. approaching those conditions thought to characterise marine calcifiers, discussed in more detail below. However, the resulting lower Mg^{2+} concentration of the seawater, and therefore the ACC precipitates,⁴¹ has been shown to reduce the time required for this ACC to transform into calcite or monohydrocalcite,⁵¹ with the implication that although the DIC required to precipitate ACC is greatly reduced, the resulting ACC is likely less stable. Indeed, large

structural differences between ACC from normal and low-Mg seawater were observed as discussed in Sec. 3.1.

In contrast to the effect of reducing $[Mg^{2+}]$, decreasing Mg/Ca_{sw} by elevating seawater $[Ca^{2+}]$ at pH 10 results in a higher ACC_{SI} (Fig. 8B). This is borne out by the experimental data overall, which broadly fall on a Mg/Ca_{sw} - ACC_{SI} array characterised by a negative slope (Fig. 9A, Tab. 2), which means that increasing Ca^{2+} does not result in a proportionally equivalent CO_3^{2-} decrease to maintain a constant solution $[Ca^{2+}][CO_3^{2-}]$ (the definition of ACC_{SI}). Nonetheless, although a higher $[Ca^{2+}]$ results in a higher ACC_{SI} , precipitation is possible at a lower DIC and $[CO_3^{2-}]$ compared to normal seawater (Fig. 8B), as in the case when Mg/Ca_{sw} is reduced by lowering $[Mg^{2+}]$. Moreover, performing a similar experiment at a lower pH of 9.5 highlights the importance of considering seawater elemental and carbonate chemistry in tandem. In contrast to the experiments at pH 10, increasing $[Ca^{2+}]$ at pH 9.5 yields precipitation at an approximately constant ACC_{SI} (Fig. 8B) and even lower DIC. The implication of this is that seawater chemistry exerts a bigger influence on the dynamics of ACC precipitation at lower pH, in addition to the wider structural variation in ACC precipitated at lower pH discussed above (Fig. 4).

In summary, we find that reducing Mg/Ca_{sw} , either by raising Ca^{2+} or lowering Mg^{2+} is the principal means by which ACC precipitation may be formed from seawater with a relatively low DIC. We stress that additional considerations like the carbonate system,^{52,53} and moreover the possible presence of a variety of organic molecules,^{7,17,54} and/or confinement¹⁸ may provide additional mechanisms by which ACC can be synthesised and stabilised, potentially independently of solution major ion chemistry. However, our results demonstrate that the concentration of both Mg^{2+} and Ca^{2+} are important in determining the precipitation and structure of ACC, and that precipitation and structure are more sensitive to solution ion chemistry at lower pH. Lastly, we find that the presence of Mg in seawater is critical to ACC precipitation. None of our experiments in Mg-free ASW yielded ACC, instead resulting in rapidly-precipitated calcite (Figs. 3F,5).

ACC precipitation at different amino acid concentrations demonstrates that the presence of all three amino acids increase ACC_{SI} . As is the case for crystalline $CaCO_3$,⁵⁵ the effect of the acidic amino acids is distinct from glycine. Specifically, we find that [Gly] exerts a relatively minor, approximately linear, control on ACC_{SI} , whilst the effect of increasing [Asp] or [Glu] is both more pronounced and nonlinear (Fig. 8E). Relatively low concentrations of both these acidic amino acids result in an increase in ACC_{SI} from ~150 to 200, i.e. greater than the change affected by the entire studied range of pH and DIC. Above an amino acid concentration of 20 mM there is little further effect, indicating that the inhibition of ACC precipitation by Asp and Glu becomes saturated above this point. As in the

case of Mg^{2+} , our results show that the amino acids, as well as playing a stabilising role,⁴⁷ also serve to inhibit ACC precipitation.

Varying the seawater carbonate system in the presence of 50 mM Asp and three different [Gly] (Fig. 8F) again highlights the importance of considering all aspects of solution chemistry when studying ACC precipitation. For example, at the lower range of [Gly] studied here (2 mM), ACC_{SI} falls on the amino acid-free variable pH/DIC array (Fig. 8F). However, at an [Asp] or [Gly] of 50 mM, the data fall above the array, as the presence of high concentrations of amino acids inhibits ACC precipitation, but crucially, the degree to which this is the case strongly depends on the seawater carbonate chemistry. At the lower pH range investigated here (~9), the inhibitory effect on ACC precipitation is large (ACC_{SI} is much higher compared to the amino acid free experiments), but at pH 10, this inhibitory effect is counteracted by the basic solution conditions such that the presence of high concentrations of both Gly and Asp do not result in a greatly increased ACC_{SI} compared to the amino acid-free experiments. The divergent response of ACC precipitation to the presence of amino acids at different seawater pH echoes the pH-dependent structural variations in ACC discussed with respect to the FTIR spectra above (Fig. 4), highlighting the importance of considering solution carbonate chemistry in concert with the Mg/Ca ratio and presence of additives.

3.3 Relevance to marine calcifiers

We were unable to precipitate ACC within the pH and DIC range thought to characterise tropical zooxanthellate corals^{26,35,36} in seawater with a Mg/Ca ratio close to natural (53/10.3 mM; Fig. 2), and the addition of amino acids serves to inhibit ACC precipitation (Fig. 8E). However, raising the seawater $[\text{Ca}^{2+}]$ or lowering $[\text{Mg}^{2+}]$ can reduce the DIC and $[\text{CO}_3^{2-}]$ necessary to precipitate ACC (Fig. 8). As such, we consider whether ACC can be produced within the DIC and pH range of marine calcifiers through a modification of the Mg/Ca ratio of the calcifying space. Fig. 10 summarises the effects of these processes in a quantitative manner, displaying the $[\text{CO}_3^{2-}]$ necessary for ACC to precipitate from seawater as a function of both seawater $[\text{Mg}^{2+}]$ and $[\text{Ca}^{2+}]$. This exercise highlights two important aspects of our dataset. Firstly, raising the initial seawater $[\text{Ca}^{2+}]$ from 10.3 to 15-50 mM results in a greatly reduced $[\text{CO}_3^{2-}]$ required for ACC precipitation, whilst the degree to which this is the case is sensitive to seawater pH. At a lower, more biologically-relevant pH of 9.5, ACC precipitation occurs at a $[\text{CO}_3^{2-}]$ ~2.5 mM lower compared to pH 10 (possibly because it differs structurally), such that the necessary $[\text{CO}_3^{2-}]$ to form ACC can be reduced to well within the range of that reconstructed for marine calcifiers. Secondly, whilst lowering the seawater $[\text{Mg}^{2+}]$ can also result in a reduction in the $[\text{CO}_3^{2-}]$ required to precipitate ACC, the extent to which this is the case is even more strongly dependent on the solution carbonate chemistry, and reducing $[\text{Mg}^{2+}]$ is a less efficient mechanism by which the necessary $[\text{CO}_3^{2-}]$ for ACC precipitation can be reduced (Fig. 10). Moreover, there is a limit to the extent to which the $[\text{CO}_3^{2-}]$ required to induce ACC precipitation can be reduced

by lowering $[\text{Mg}^{2+}]$. The experiments in seawater with 10 mM Mg^{2+} and DIC at the lower end of the range investigated likely represent ACC-calcite mixtures (Fig. 7D), and further decreasing the initial seawater Mg/Ca ratio resulted in calcite precipitation.

In summary, these results indicate that manipulation of the calcification site Mg/Ca ratio may enable ACC to be precipitated at a relatively low DIC. However, applying this finding to the tropical zooxanthellate corals results in a discrepancy. Although the presence of an ACC precursor phase has been reported for these organisms,^{10,56} several studies using independent techniques have shown that the $[\text{Ca}^{2+}]$ at the calcification site of corals is unlikely to be elevated above ~15 mM.^{26,43,57} ACC precipitation at this $[\text{Ca}^{2+}]$ required a $[\text{CO}_3^{2-}]$ of >8 mM in our experiments (Fig. 10), far exceeding that thought to occur at the biomineralization site of these corals.^{26,28,43,57,58} We also note that the experimental transformation of inorganic ACC to aragonite is a long process, taking ~10 days in seawater with the normal Mg/Ca ratio and proceeding through an intermediate monohydrocalcite phase.¹⁹ Lastly, it may be difficult to reconcile observed skeletal trace element distribution coefficients with a greatly elevated $[\text{Ca}^{2+}]$.²⁴ It is beyond the scope of this study to address this issue directly, except to highlight that reconciliation of our precipitation data with that from corals either requires the role of ACC in corals to be reconsidered, or for a mechanism to be found by which the DIC necessary to precipitate ACC can be reduced without greatly modifying the seawater Mg/Ca ratio. Irrespective, one specific area in which our results can directly inform the ongoing debate regarding ACC as a possible precursor to coral aragonite (cf. refs. 59,60) concerns the interpretation of Raman spectra. A distinguishing feature of ACC is the greatly increased ν_1 FWHM relative to crystalline CaCO_3 and lack of visible lattice mode vibrations (Fig. 7). Pertinently, we show that material precipitated under certain conditions has spectral features of both ACC and calcite (Fig. 3E), and likely represents ACC-calcite mixtures (Fig. 7D). Care should be taken in the identification of biological ACC based on Raman spectra that contain lattice mode vibrations. Whilst lattice mode vibrations do not necessarily rule out the presence of an amorphous phase, the presence or position of lattice mode vibrations alone are not indicative of ACC.

In contrast to marine calcifiers that build an aragonitic skeleton, our data are not necessarily difficult to reconcile with biomineralization in organisms that produce a low-Mg calcite shell such as the planktonic foraminifera. These organisms have also been suggested to calcify through an ACC precursor,^{11,12} but crucially there is consensus that the Mg/Ca ratio at the site of calcification must be greatly reduced below that of seawater in order to promote the formation of low-Mg calcite.^{13,30,61} However, at present little is known about the calcification site DIC in foraminifera. A more detailed comparison to our inorganic ACC precipitation data will require better constraint of this aspect of foraminifera calcification.

4. Conclusion

The control that pH and DIC, the concentration of $[\text{Mg}^{2+}]$ and $[\text{Ca}^{2+}]$, and the presence of different concentrations of Asp, Glu and Gly exert on the saturation state required to precipitate ACC from seawater (ACC_{SI}) was investigated. In normal seawater ($\text{Mg}/\text{Ca} = 5$), the solution pH and DIC exert at most a minor control on ACC_{SI} and no resolvable structural control as evidenced by FTIR, Raman, XRD, and TGA-MS. The presence of high concentrations (>2 mM) of amino acids inhibit ACC precipitation, but also do not alter the spectroscopic features or water of crystallisation of the precipitate. In contrast, lowering the seawater Mg/Ca ratio, either by reducing $[\text{Mg}^{2+}]$ or increasing $[\text{Ca}^{2+}]$, may greatly reduce the DIC and $[\text{CO}_3^{2-}]$ necessary to precipitate, yet can result in a greater degree of short and long-range order, as evidenced by the appearance of lattice-mode vibrations, a narrower ν_3 Raman peak, and a greatly reduced water of crystallisation.

In addition, we investigate the combined effect of varying the seawater carbonate chemistry and $[\text{Mg}^{2+}]$, $[\text{Ca}^{2+}]$ or amino acid concentration on the structure of ACC and ACC_{SI} . Whilst we find no substantial direct impact of pH or DIC on ACC structure precipitated from unmodified seawater (see above), these experiments highlight the differing impact of the seawater Mg/Ca ratio and concentration of amino acids at lower pH. In general, modifying $\text{Mg}/\text{Ca}_{\text{sw}}$ exerts a much greater control on both the structure of ACC and ACC_{SI} at pH 9-9.6 compared to pH 10, the latter typical of much inorganic ACC work, highlighting the need to consider the solution carbonate chemistry in tandem with other variables.

Finally, we find that the DIC required to precipitate ACC from normal seawater greatly exceeds that thought to characterise the calcifying space of certain marine organisms that may utilise this pathway. The only variable investigated here which resulted in a large, systematic decrease in the DIC required to precipitate ACC is the seawater Mg/Ca ratio, raising the possibility that modifying the seawater Mg/Ca ratio through Mg removal or Ca addition is an important component of utilising an ACC pathway.

Supporting Information. Further methodological details, additional TGA-MS results, and an in-depth assessment of the multiple linear regression models relating ACC_{SI} to the experimental variables. Supplementary Tab. 1 presents full seawater chemistry details for all precipitates.

Acknowledgement

This work was supported by the Leverhulme Trust (Research project grant 2015-268 to NA, RK, and KP). The Royal Society is gratefully acknowledged for the award of an Industry Fellowship to P.B.W. We are thankful to Laetitia Pichevin for assistance in analysing seawater samples by ICP-OES.

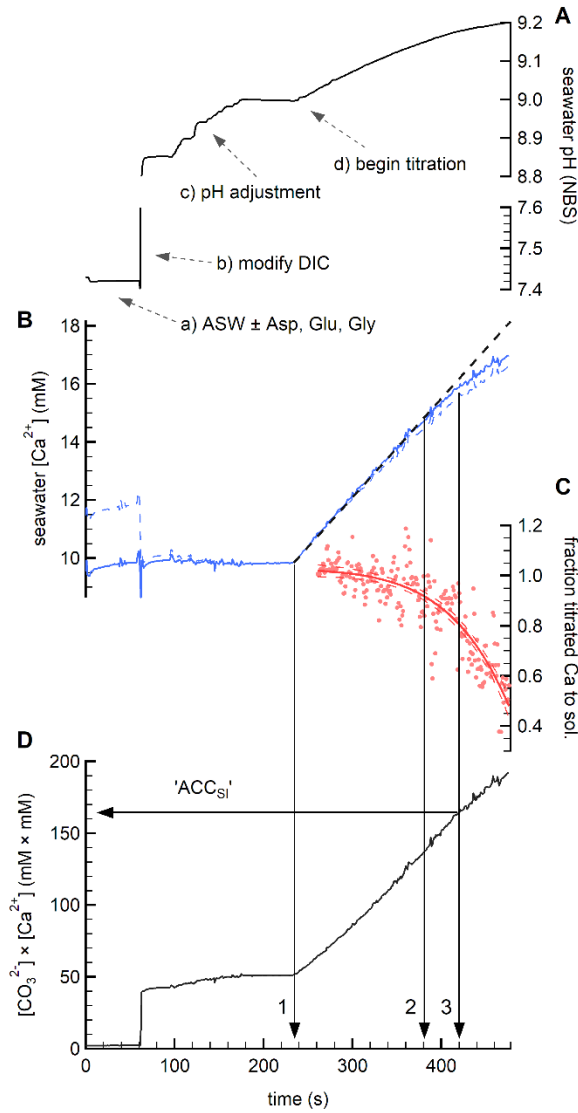


Figure 1: An example titration curve to induce ACC precipitation from seawater. (A) pH evolution over the course of the titration, note break in y-axis scale. In this case, (i) artificial seawater is (ii) spiked with 1 M Na_2CO_3 to achieve the desired DIC for the experiment. (iii) The pH is then adjusted using HCl or NaOH, and the solution is equilibrated for 60 s. (iv) ACC precipitation is induced by simultaneous titration of 0.45 M Na_2CO_3 and CaCl_2 . A pH increase occurs as ACC precipitation is not immediate. (B) Seawater $[\text{Ca}^{2+}]$ measured in real-time using a calibrated ISE. Dashed and solid blue lines show the measurement both uncorrected and corrected for pH change, as pH exerts a minor control on the measured electrode voltage. The dashed black line shows the predicted evolution of seawater Ca^{2+} if no precipitation takes place. The divergence of the Ca electrode measurement from this line therefore indicates the onset of precipitation. (C) The fraction Ca titrated that went into solution, based on the divergence of the curves shown in panel B. Vertical arrows (1) show the onset of titration, (2) the divergence of the measured solution Ca^{2+} from predicted, indicating the initiation of precipitation, and (3) the point at which more than 20% of the titrated Ca was utilised in ACC precipitation. (D) The temporal evolution of the product of $[\text{Ca}^{2+}]$ and $[\text{CO}_3^{2-}]$, the latter calculated from DIC and pH using co2sys.³⁶ In the text and subsequent figures the seawater saturation state required to precipitate ACC (abbreviated ‘ ACC_{SI} ’) is calculated as being the $[\text{Ca}^{2+}] \cdot [\text{CO}_3^{2-}]$ product at the location of arrow 3.

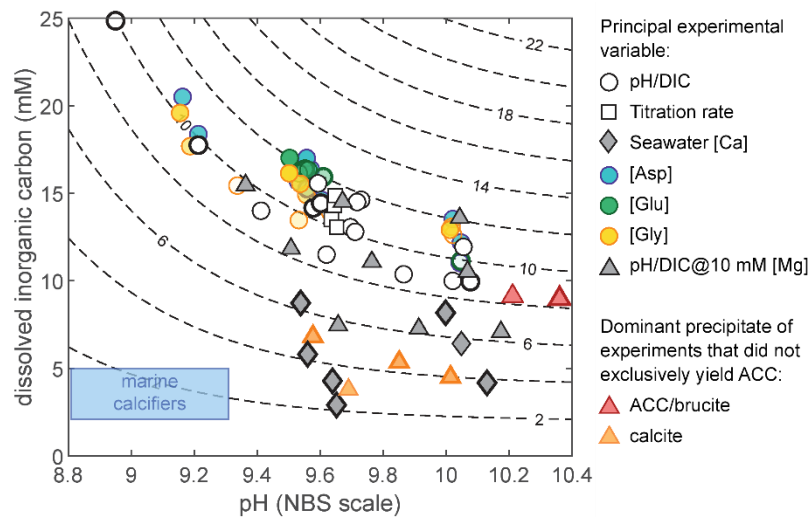


Figure 2: The pH and DIC range over which ACC was precipitated from seawater in this study. DIC and pH broadly co-varied in our experiments as it was not possible to precipitate ACC from (e.g.) low-pH, low-DIC seawater without modifying the seawater Mg/Ca ratio. The approximate upper range of estimates of carbonate chemistry at the site of biomineralisation in tropical zooxanthellate corals and foraminifera is shown for comparison (transparent blue box, note that the calcification site DIC estimates relate only to corals).^{27,28,36} Contours show lines of equal $[\text{CO}_3^{2-}]$ in mM. In most cases ACC was precipitated from solutions with a $[\text{CO}_3^{2-}]$ of 10 ± 2 mM, approximately equivalent to a seawater $\text{Ca}^{2+}/\text{CO}_3^{2-}$ ratio of 1. Amino acid concentration is shown as a function of opacity, lighter symbols indicate lower concentrations. Bold symbols indicate experiments for which solution chemistry was directly measured, others are calculated following the method outlined in Fig. 1.

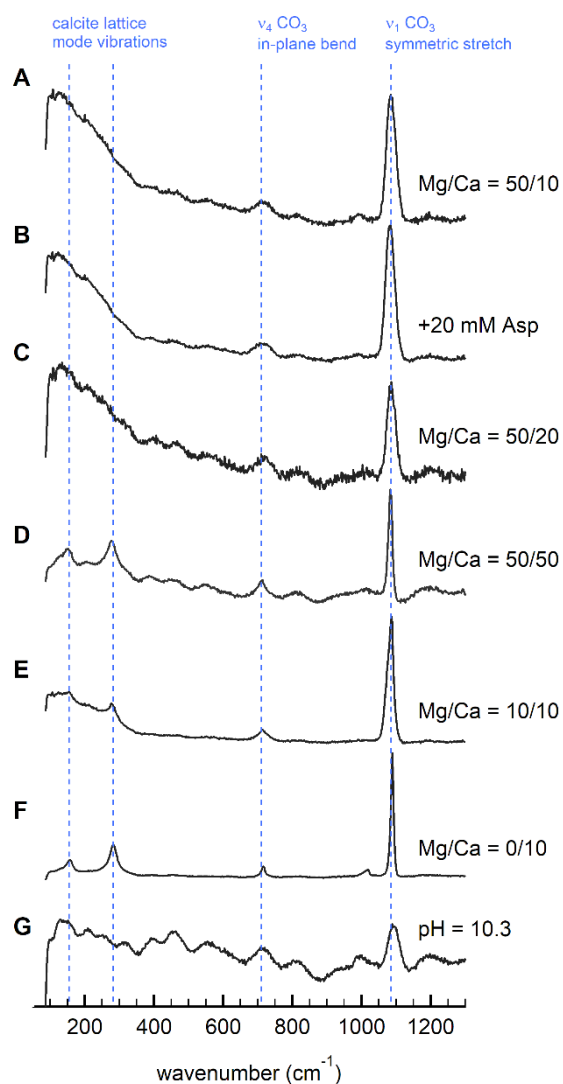


Figure 3. Representative Raman spectra of the inorganic precipitates. No processing was applied except for a linear baseline correction. Mg/Ca labels give the concentration of both elements in mM. All spectra show the characteristic features of ACC, except for F (calcite), and G, precipitated under extreme basic conditions (pH 10.3). At low Mg/Ca_{sw} ratios the lattice mode vibrations of calcite become visible (D, E).

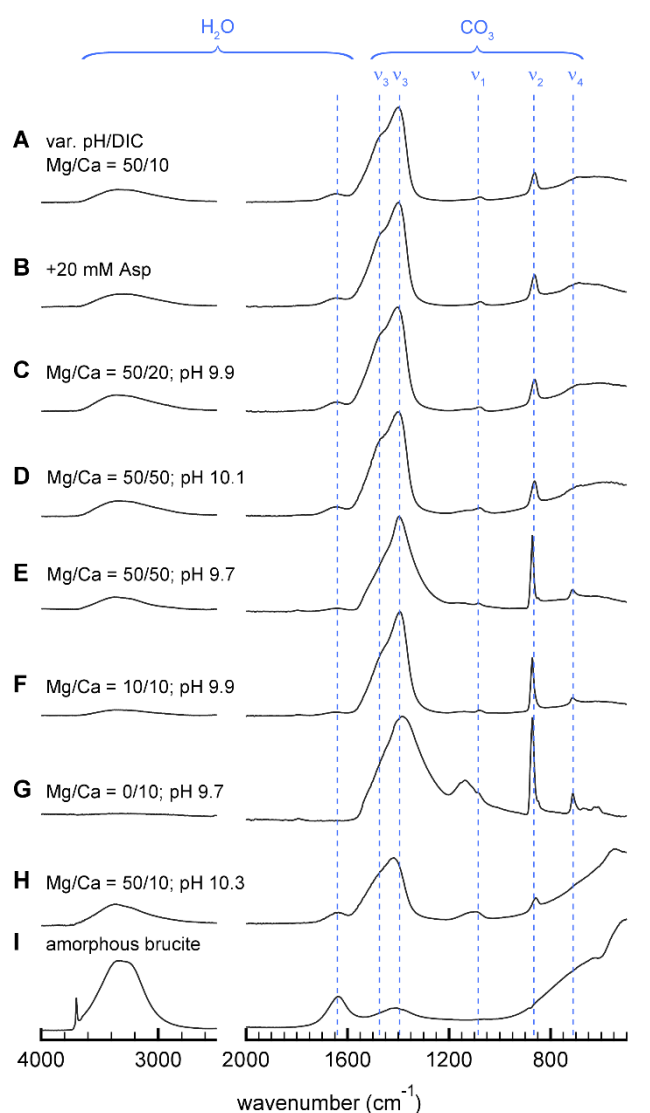


Figure 4: Representative baseline-corrected ATR-FTIR spectra of the precipitates. Mg/Ca labels give the concentration of both elements in mM. Spectra A-F are of ACCs, which have a characteristic split ν_3 vibration at 1410 and 1474 cm^{-1} , G is calcite, and H, precipitated under extreme basic conditions, is an ACC-amorphous brucite mixture. Amorphous hydrated brucite is shown for comparison in I, prepared by pipetting 1 M MgCl_2 into deionised water at pH 11, achieved through NaOH addition. Note the structural differences between D, E and F, precipitated under the same seawater Mg/Ca ratio, but at different pH and absolute $[\text{Mg}^{2+}]$ and $[\text{Ca}^{2+}]$. The peak at 1648 cm^{-1} is an OH vibration,¹⁶ and does not indicate a brucite component in the ACC samples.

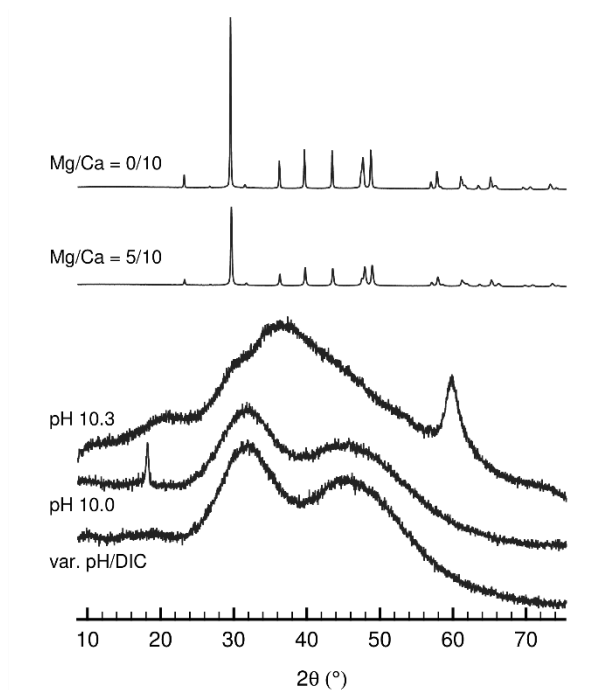


Figure 5: Representative XRD spectra of selected precipitates. A characteristic spectrum from the variable pH/DIC experiment is shown at the bottom of the figure. The broadly featureless spectrum confirms the amorphous or nanocrystalline nature of these samples. The only deviations from this spectrum were those of precipitates formed at high pH, which contain small, unidentifiable peaks, and at low seawater $[\text{Mg}^{2+}]$ (<10 mM), which are partly or entirely calcite.

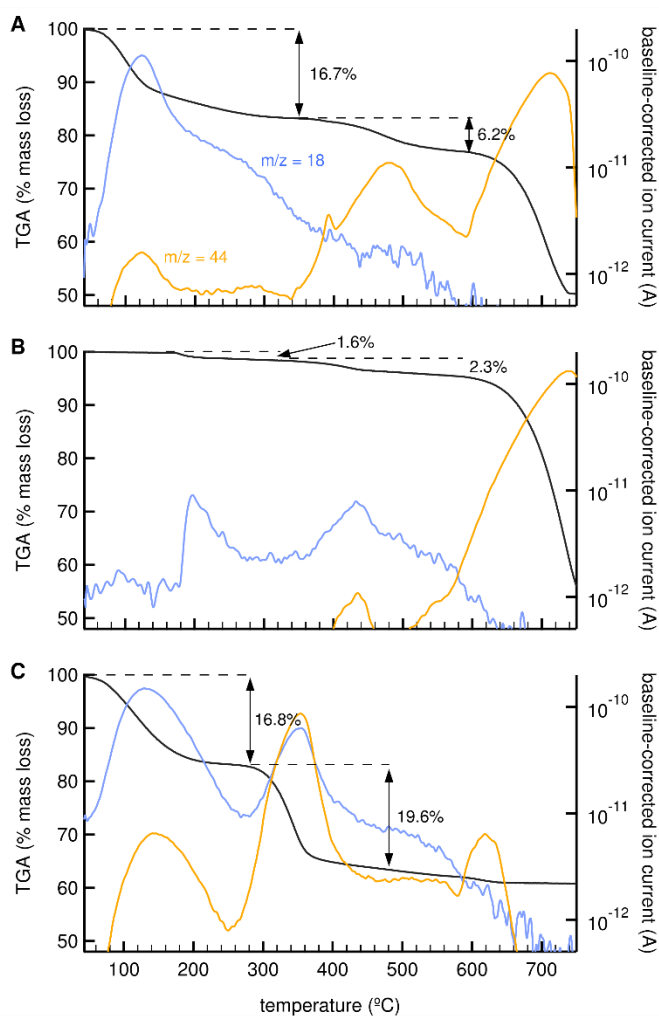


Figure 6: TGA-MS profiles of three endmember precipitates. Products of decomposition are shown in blue (H₂O, $m/z = 18$) and yellow (CO₂, $m/z = 44$). (A) ACC precipitated in seawater at pH 8.95 with al Mg/Ca ratio close to natural ($\sim 5 \text{ mol mol}^{-1}$), showing the characteristic three-stage decomposition of these inorganic ACCs. (B) Calcite precipitation from seawater, achieved by reducing the seawater [Mg²⁺] by a factor of 10 compared to natural (5 mM). (C) Amorphous material precipitated at the basic extreme of the pH range investigated (10.3 on the NBS scale). The FTIR spectra of this material indicates that it is an amorphous CaCO₃-brucite mixture.

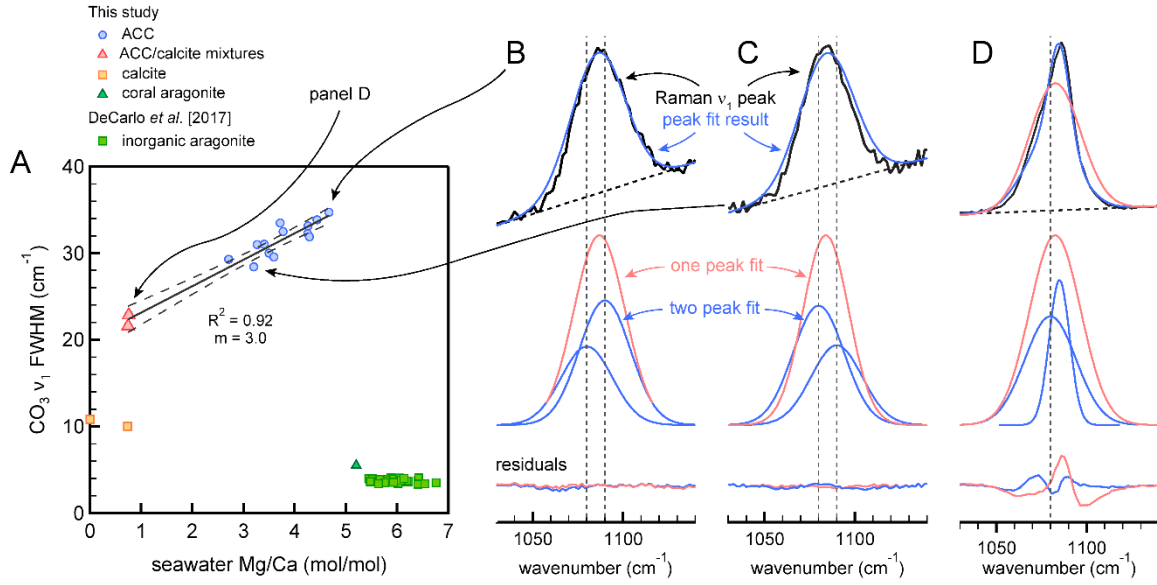


Figure 7: (A) Solution chemistry control on the Raman ν_1 (CO_3 symmetric stretch) FWHM at $\sim 1080 \text{ cm}^{-1}$ of ACCs precipitated from seawater, compared to calcite precipitated under similar carbonate chemistry conditions (see the Supplementary Information), inorganic aragonite,⁴³ and a natural coral aragonite (*Porites* spp.). The latter is plotted assuming that corals precipitate aragonite from seawater with an unmodified Mg/Ca ratio, which may not be the case.²⁶ (B-D) Gaussian peak fitting of the ν_1 band of three selected amorphous precipitates spanning the investigated range of seawater Mg/Ca ratios. Panels B and C represent samples precipitated under different $[\text{Ca}^{2+}_{\text{sw}}]$, whereas D was precipitated with $\text{Mg}^{2+}_{\text{sw}}$ reduced from 53 to 10 mM. The ν_1 band in ACCs precipitated under variable $[\text{Ca}^{2+}_{\text{sw}}]$ is symmetric (these spectra can be fit equally well with one or two peaks). In contrast, ACC precipitated at the lowest seawater Mg/Ca ratio is characterised by an asymmetric ν_1 band which is best modelled by two peaks with different widths. This suggests that these precipitates are physical mixtures of low-Mg ACC and calcite.

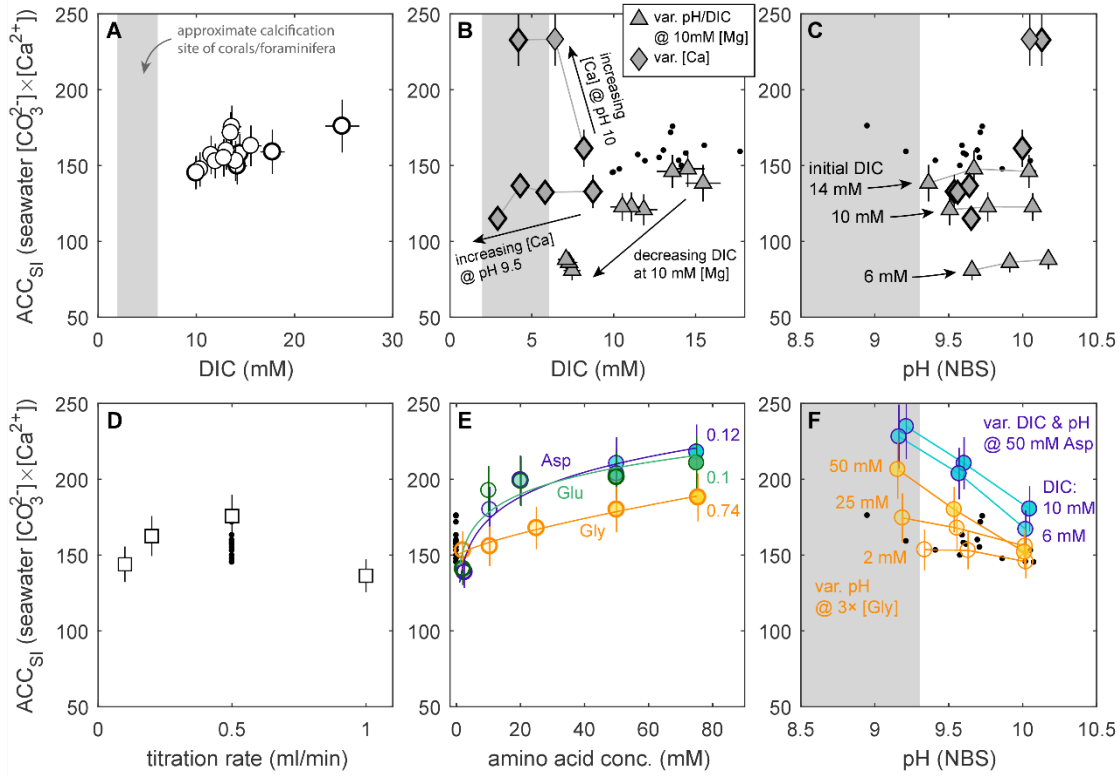


Figure 8: The control exerted by solution carbonate chemistry, Mg/Ca ratio and amino acid concentration on ACC_{SI}, the seawater saturation state necessary to precipitate ACC. Bold symbols are experiments for which solution chemistry was directly measured, others are calculated following the method outlined in Fig. 1. Vertical grey bars delineate the approximate range of DIC and pH at the site of biomineralisation in tropical zooxanthellate corals and foraminifera.^{21,25} (A) ACC_{SI} as a function of DIC. Only the amino acid-free experiments in normal seawater (Mg/Ca = ~5) are shown. (B) The differential response of ACC_{SI} to reducing Mg/Ca_{sw} through varying seawater [Mg²⁺] and [Ca²⁺]. Both adjustments result in ACC precipitation at lower DIC compared to the experiments in normal seawater (black dots from panel A). (C) The effect of varying pH and DIC in seawater with a reduced Mg/Ca ratio (10 mM Mg²⁺ instead of 53 mM or 20-50 mM Ca²⁺ instead of 10 mM, see panel B for legend). Lines join experiments of equivalent initial DIC at different pH. Note that the low DIC experiments may contain a calcite component (Fig. 7D). (D) Titration rate exerts no systematic effect on ACC_{SI}. (E) The impact of amino acid concentration on ACC precipitation. Power coefficients of least-squares regressions are shown. (F) The combined influence of varying pH/DIC in the presence of Asp and Gly. Low AA concentrations (2 mM) produce data that fall within the range of the additive-free experiments (black dots from panel A), higher concentrations strongly inhibit ACC precipitation at lower, more biologically relevant pH, but to a much lesser extent at high pH. Amino acid concentration is shown as a function of symbol opacity in panels E and F.

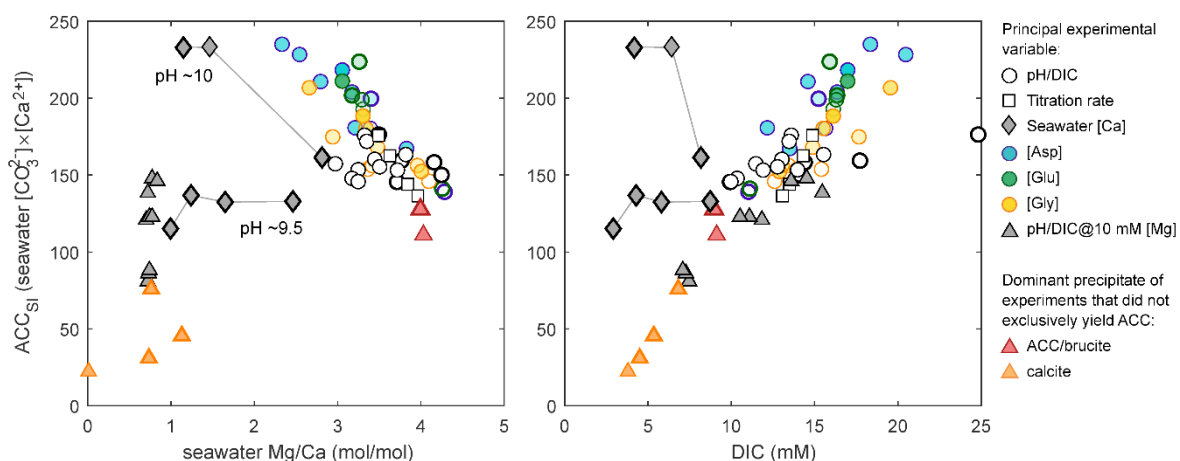


Figure 9: Combining the data shown in Fig. 8 to examine the overall controls of solution chemistry on ACC_{SI} , the seawater saturation state required to induce ACC precipitation. Most experiments, especially those performed in the presence of amino acids, fall on a negative ACC_{SI} -Mg/Ca_{sw} array and positive ACC_{SI} -DIC array, indicating that the inhibitive effect of these additives is overcome through Ca^{2+} and/or DIC titration (see also the multiple linear regression models in Tab. 2). Error bars are omitted for clarity (see Fig. 8).

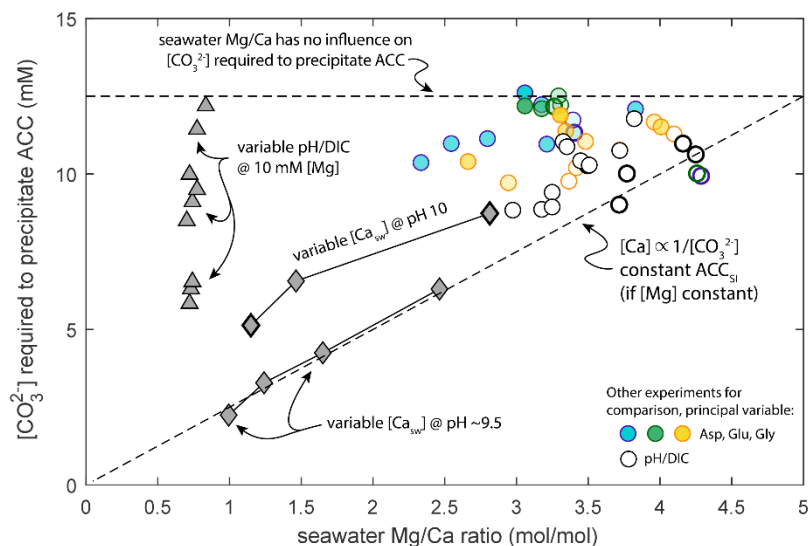


Figure 10: Comparing strategies to reduce the $[\text{CO}_3^{2-}]$ necessary for ACC precipitation from seawater; Mg^{2+} removal (triangles) or Ca^{2+} addition (diamonds). Data from experiments in which $\text{Mg}/\text{Ca}_{\text{sw}}$ was not the principal variable are shown for comparison (circles). Dashed lines show hypothetical endmember scenarios of the relationship between ACC precipitation and solution chemistry. Almost no data lie on the horizontal dashed line, with the exception of experiments that contained high concentrations of amino acids, demonstrating that reducing $\text{Mg}/\text{Ca}_{\text{sw}}$ either by raising the seawater $[\text{Ca}^{2+}]$ or reducing $[\text{Mg}^{2+}]$ lowers the $[\text{CO}_3^{2-}]$ necessary for ACC precipitation. In addition, these data highlight the interplay of solution elemental and carbonate chemistry. At a higher (less biologically relevant) pH, raising $[\text{Ca}^{2+}_{\text{sw}}]$ results in a reduction in the required $[\text{CO}_3^{2-}]$ required to precipitate ACC, but to a lesser extent than at pH 9.5, demonstrating that ACC precipitation is differently (more) sensitive to $[\text{Ca}^{2+}_{\text{sw}}]$ at lower pH.

Table 1: Overview of the investigated factors impacting ACC precipitation from seawater. The carbonate chemistry details represent those at the onset of precipitation (i.e. at ACC_{SI}), whereas the seawater [Mg²⁺] and [Ca²⁺] are those at the start of the experiment. See the Supplementary Information for full details.

Principal experimental variable	n	DIC (mM) at ACC _{SI}	pH (NBS) at ACC _{SI}	Titration rate (ml/min)	Amino acid conc. (mM)	Initial seawater [Mg] (mM)	Initial seawater [Ca] (mM)	Detailed sample ID (see SI)
pH & DIC	14	10.0-24.9	9.0-10.1	0.5	-	53	10.3	1-1, 1-2, 1-3, 1-13
Titration rate	4	13.0-14.9	9.6	0.1-1.0	-	53	10.3	1-15, 1-16
pH & DIC in low-Mg (10 mM) seawater	3×3	7.1-15.5	9.4-10.2	0.5	-	10	10.3	1-17
[Ca] @ 2× pH	3	3.0-8.7	9.5-9.7	0.5	-	53	15.0-45.0	1-6, 1-7
	4	4.2-8.2	10.0-10.1				20.0-50.0	2-19
pH & DIC with 50 mM Asp	6	12.2-20.5	9.2-10.0	0.5	50	53	10.3	-
[Asp]	5	11.0-17.0	9.5-10.0	0.5	2.5-75	53	10.3	1-4, 1-10
[Glu]	5	11.1-17.0	9.5-10.0	0.5	2-75	53	10.3	1-5, 1-11
[Gly] @ 3×pH	11	12.6-19.6	9.2-10.0	0.5	2-75	53	10.3	-
[Mg] & [Ca] @ Constant Mg/Ca	2	3.0,14.5	9.7	0.5	-	53,10	50,10.3	-
Mg-free SW	1	3.8	9.7	0.5	-	0.1	10.3	1-8
High pH extreme	2	9.0-9.1	10.2-10.4	0.5	-	53	10.3	1-9

Table 2: Gradients and goodness of fit of multiple linear regression models through subsets of the ACC precipitation data shown in Fig. 2 and summarised in Tab. 1. The models were produced twice, both with and without amino acid concentration as an independent variable, otherwise taking the form defined in Eq. 2 (see text for rationale). The variable pH/DIC model was also performed twice, both with and without one outlier removed (the highest DIC data point in Fig. 8A, based on its Cook's distance). Coefficient uncertainties are 1SE except where noted. Bold values denote independent variables that are significant predictors of ACC_{SI} at the 95% CI. Note that a coefficient p value >0.05 does not necessarily imply that the predictor is not significant, as this can also arise from covariance in the predictors (e.g. amino acid concentration and Mg/Ca_{sw}; Fig. 9). The units of the slopes for Mg/Ca_{sw}, pH, DIC, and [AA] are mol/mol⁻¹, unit⁻¹, mM⁻¹, and mM⁻¹ respectively.

Principal variable:	pH/DIC		pH/DIC		Aspartic acid conc.		Glutamic acid conc. [‡]		Glycine conc.		pH/DIC in low-Mg (10 mM) seawater		Entire dataset [†]	
	m*	p*	m*	P*	m	p	m	p	m	p	m	p	m	p
Amino acid concentration used in multiple linear regression model														
Mg/Ca _{sw}	-9.4±5.6	0.15	-16.2±4.0	0.01	-10.4±9.6	0.33	-9.1		-40.0±23.6	0.14	16.5±106.1	0.882	-30.0±4.6	4.3×10 ⁻⁷
pH	24.3±16.6	0.14	37.6±11.8	0.01	-96.1±35.5	0.04	74.4		40.7±25.6	0.16	25.3±16.9	0.195	47.5±14.2	2.0×10 ⁻³
DIC	4.0±1.2	0.01	7.1±1.4	5×10 ⁻⁴	-4.9±3.3	0.19	16.2		4.0±2.7	0.19	8.8±1.3	9.1×10 ⁻⁴	8.7±1.6	3.5×10 ⁻⁶
[AA]	-	-	-	-	0.6±0.2	0.02	0.1		0.2±0.1	0.06	-	-	0.17±0.09	0.06
Overall model:														
R ²	0.66		0.79		0.98		1.00		0.95		0.98		0.89	
p	0.01		2.0×10 ⁻³		1.9×10 ⁻⁴				4.5×10 ⁻⁴		1.8×10 ⁻⁴		1.3×10 ⁻¹⁵	
RMSE	6.9		5.2		5.6				5.4		5.1		9.3	
Amino acid concentration not used in multiple linear regression model														
Mg/Ca _{sw}	As above		As above		-35.8±9.1	7.5×10 ⁻³	-30.9±9.5	0.19	-62.8±27.3	0.05	As above		-33.5±4.2	2.1×10 ⁻⁹
pH					1.6±30.5	0.96	101.2±21.1	0.13	77.5±25.6	0.02			64.0±11.7	4.0×10 ⁻⁶
DIC					3.5±3.3	0.34	14.9±3.3	0.82	5.3±3.4	0.16			10.7±1.2	2.6×10 ⁻¹⁰
Overall model:														
R ²					0.94		1.00		0.91				0.88	
p					4.9×10 ⁻⁴		0.02		5.4×10 ⁻⁴				7.6×10 ⁻¹⁶	
RMSE					9.0		0.8		6.8				9.7	

*These estimates represent the median±2SD values of a Monte Carlo simulation (1000 models) including normally-distributed uncertainty in both dependent and independent variables. For all other models the least-squares regression through the best estimate of all data are shown. See the SI for a demonstration that there is no significant difference between the two approaches.

‡There are insufficient data from this subset of experiments to reliably estimate coefficient or model uncertainties for this number of independent variables.

†The experiments conducted in low-Mg seawater were excluded from this regression model as some precipitates are likely ACC-calcite mixtures (see text). The model that uses amino acid concentration as a predictor considers all amino acids together, i.e. Asp, Glu, and Gly were not included as separate terms. This choice makes little difference to the result (see the SI).

References

- (1) Hodson, M. E.; Benning, L. G.; Demarchi, B.; Penkman, K. E. H.; Rodriguez-Blanco, J. D.; Schofield, P. F.; Versteegh, E. A. A. Biomineralisation by earthworms – an investigation into the stability and distribution of amorphous calcium carbonate *Geochem. Trans.* **2015**, *16* (4), doi:10.1186/s12932-015-0019-z.
- (2) Addadi, L.; Raz, S.; Weiner, S. Taking advantage of disorder: Amorphous calcium carbonate and its roles in biomineralization *Adv. Mater.* **2003**, *15*, 959–970.
- (3) Taylor, M. G.; Simkiss, K.; Greaves, G. N.; Okazaki, M.; Mann, S. An X-ray absorption spectroscopy study of the structure and transformation of amorphous calcium carbonate from plant cystoliths *Proc. R. Soc. B Biol. Sci.* **1993**, *252*, 75–80.
- (4) Weiner, S.; Levi-Kalishman, Y.; Raz, S.; Addadi, L. Biologically formed amorphous calcium carbonate *Connect. Tissue Res.* **2003**, *44*, 214–218.
- (5) Aizenberg, B. J.; Lambert, G.; Addadi, L.; Weiner, S. Stabilization of amorphous calcium carbonate by specialized macromolecules in biological and synthetic precipitates *Adv. Mater.* **1996**, *8*, 222–226.
- (6) Beniash, E.; Aizenberg, J.; Addadi, L.; Weiner, S. Amorphous calcium carbonate transforms into calcite during sea urchin larval spicule growth Amorphous calcium carbonate transforms into calcite during sea urchin larval spicule growth *Proc. R. Soc. B Biol. Sci.* **1997**, *264*, 461–465.
- (7) Aizenberg, J.; Lambert, G.; Weiner, S.; Addadi, L. Factors Involved in the formation of amorphous and crystalline calcium carbonate : A study of an Ascidian skeleton *J. Am. Chem. Soc.* **2002**, *124*, 32–39.
- (8) Weiss, I. M.; Tuross, N.; Addadi, L.; Weiner, S. Mollusc larval shell formation: Amorphous calcium carbonate is a precursor phase for aragonite *J. Exp. Zool.* **2002**, *293*, 478–491.
- (9) Sviben, S.; Gal, A.; Hood, M. A.; Bertinetti, L.; Politi, Y.; Bennet, M.; Krishnamoorthy, P.; Schertel, A.; Wirth, R.; Sorrentino, A.; Pereiro, E.; Faivre, D.; Scheffel, A. A vacuole-like compartment concentrates a disordered calcium phase in a key coccolithophorid alga *Nat. Commun.* **2016**, *7*, doi:10.1038/ncomms11228.
- (10) Mass, T.; Giuffrè, A. J.; Sun, C.; Stiffler, C. A.; Frazier, M. J.; Neder, M.; Tamura, N.; Stan C. V.; Marcus, M. A.; Gilbert, P. U. P. A. *Proc. Natl. Acad. Sci.* **2017**, *114*, E7670–E7678.
- (11) Erez, J. The source of ions for biomineralization in foraminifera and their implications for paleoceanographic proxies *Rev. Miner. Geochem.* **2003**, *54*, 115–149.
- (12) Jacob, D. E.; Wirth, R.; Agbaje, O. B. A.; Branson, O.; Eggins, S. M. Planktic foraminifera form their shells via metastable carbonate phases *Nat. Commun.* **2017**, *8*, 1265.
- (13) Evans, D.; Müller, W.; Erez, J. Assessing foraminifera biomineralisation models through trace element data of cultures under variable seawater chemistry *Geochim. Cosmochim. Acta* **2018**, *236*, 198–217.
- (14) Schiebel, R. Planktic foraminiferal sedimentation and the marine calcite budget *Glob. Biogeochem. Cycles* **2002**, *16*, doi:10.1029/2001GB001459.
- (15) Henahan, M. J.; Evans, D.; Shankle, M.; Burke, J. E.; Foster, G. L.; Anagnostou, E.; Chalk, T. B.; Stewart, J. A.; Alt, C. H. S.; Durrant, J.; Hull, P. M. Size-dependent response of foraminiferal calcification to seawater carbonate chemistry *Biogeosciences* **2017**, *14*, 3287–3308.
- (16) Loste, E.; Wilson, R. M.; Seshadri, R.; Meldrum, F. C. The role of magnesium in stabilising amorphous calcium carbonate and controlling calcite morphologies *J. Cryst. Growth* **2003**, *254*, 206–218.
- (17) Bentov, S.; Weil, S.; Glazer, L.; Sagi, A.; Berman, A. Stabilization of amorphous calcium carbonate by phosphate rich organic matrix proteins and by single phosphoamino acids *J. Struct. Biology* **2010**, *171*, 207–215.
- (18) Stephens, C. J.; Ladden, S. F.; Meldrum, F. C.; Christenson, H. K. Amorphous calcium carbonate is stabilized in confinement *Adv. Funct. Mater.* **2010**, *20*, 2108–2115.
- (19) Zhang, Z.; Xie, Y.; Xu, X.; Pan, H.; Tang, R. Transformation of amorphous calcium carbonate into aragonite *J. Cryst. Growth* **2012**, *343*, 62–67.
- (20) Bots, P.; Benning, L. G.; Rodriguez-Blanco, J.-D.; Roncal-Herrero, T.; Shaw, S. Mechanistic insights into the crystallization of amorphous calcium carbonate (ACC) *Cryst. Growth Des.* **2012**, *12*, 3806–3814.
- (21) Lam, R. S. K.; Charnock, J. M.; Lennie, A.; Meldrum, F. C. Synthesis-dependant structural variations in amorphous calcium carbonate *Cryst. Eng. Comm.* **2007**, *9*, 1226–1236.
- (22) Levi-Kalishman, B. Y.; Raz, S.; Weiner, S.; Addadi, L.; Sagi, I. Structural differences between biogenic amorphous calcium carbonate phases using X-ray absorption spectroscopy *Adv. Funct. Mater.* **2002**, *12*, 43–48 (2002).
- (23) Bentov, S.; Brownlee, C.; Erez, J. The role of seawater endocytosis in the biomineralization process in calcareous foraminifera *Proc. Natl. Acad. Sci.* **2009**, *106*, 21500–21504.
- (24) Gagnon, A. C.; Adkins, J. F.; Erez, J. Seawater transport during coral biomineralization *Earth Planet. Sci. Lett.* **2012**, *329*, 150–161.
- (25) Tambutti, E.; Tambutti, S.; Segonds, N.; Zoccola, D.; Venn, A.; Erez, J.; Allemand, D. Calcein labelling and electrophysiology : insights on coral tissue permeability and calcification *Proc. R. Soc. B Biol. Sci.* **2012**, *279*, 19–27.
- (26) Al-horani, F. A.; Al-moghrabi, S. M.; de Beer, D. Microsensor study of photosynthesis and calcification in the scleractinian coral , *Galaxea fascicularis* : active internal carbon cycle *J. Exp. Marine Biol. Ecol.* **2003**,

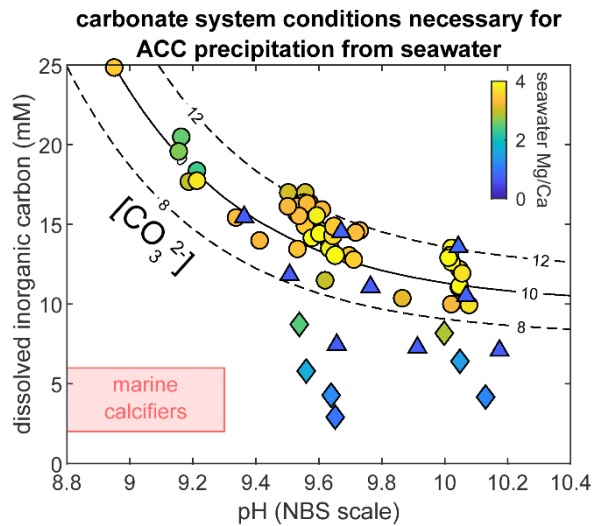
288, 1–15.

- (27) de Nooijer, L. J.; Toyofuku, T.; Kitazato, H. Foraminifera promote calcification by elevating their intracellular pH *Proc. Natl. Acad. Sci.* **2009**, *106*, 15374–15378.
- (28) Allison, N.; Cohen, I.; Finch, A.; Erez, J.; Tudhope, A. W. Corals concentrate dissolved inorganic carbon to facilitate calcification *Nat. Commun.* **2014**, *5*, doi:10.1038/ncomms6741.
- (29) Gaetani, G. A.; Cohen, A. L. Element partitioning during precipitation of aragonite from seawater : A framework for understanding paleoproxies *Geochim. Cosmochim. Acta* **2006**, *70*, 4617–4634.
- (30) Bentov, S.; Erez, J. Impact of biomineralization processes on the Mg content of foraminiferal shells: A biological perspective *Geochemistry Geophys. Geosystems* **2006**, *7*, doi:10.1029/2005GC001015.
- (31) Inoue, M.; Gussone, N.; Koga, Y.; Iwase, A.; Suzuki, A.; Sakai, K.; Kawahata, H. Controlling factors of Ca isotope fractionation in scleractinian corals evaluated by temperature, pH and light controlled culture experiments *Geochim. Cosmochim. Acta* **2015**, *167*, 80–92.
- (32) Giuffrè, A. J.; Gagnon, A. C.; De Yoreo, J. J.; Dove, P. M. Isotopic tracer evidence for the amorphous calcium carbonate to calcite transformation by dissolution-reprecipitation *Geochim. Cosmochim. Acta* **2015**, *165*, 407–417.
- (33) Faatz, B. M.; Gröhn, F.; Wegner, G. Amorphous calcium carbonate: Synthesis and potential intermediate in biomineralization *Adv. Funct. Mater.* **2004**, *16*, 996–1000.
- (34) Ogino, T.; Suzuki, T.; Sawada, K. The formation and transformation mechanism of calcium carbonate in water *Geochim. Cosmochim. Acta* **1987**, *51*, 2757–2767.
- (35) Cai, W. J.; Ma, Y.; Hopkinson, B. M.; Grottoli, A. G.; Warner, M. E.; Ding, Q.; Hu, X.; Yuan, X.; Schoepf, V.; Xu, H.; Han, C.; Melman, T. F.; Hoadley, K. D.; Pettay, D. T.; Matsui, Y.; Baumann, J. H.; Levas, S.; Ying, Y.; Wang, Y. Microelectrode characterization of coral daytime interior pH and carbonate chemistry *Nat. Commun* **2016**, *7*, doi:10.1038/ncomms11144.
- (36) McCulloch, M. T.; D’Olivo Cordero, J. P.; Falter, J.; Holcomb, M.; Trotter, J. A. Coral calcification in a changing World and the interactive dynamics of pH and DIC upregulation *Nat. Commun.* **2017**, *8*, doi:10.1038/ncomms15686.
- (37) Wang, D.; Wallace, A. F.; De Yoreo, J. J.; Dove, P. M. Carboxylated molecules regulate magnesium content of amorphous calcium carbonates during calcification *Proc. Natl. Acad. Sci.* **2009**, *106*, 21511–21516.
- (38) Whittaker, M. L.; Sun, W.; DeRocher, K. A.; Jayaraman, S.; Ceder, G.; Joester, D. Structural basis for metastability in amorphous calcium barium carbonate (ACBC) *Adv. Funct. Mater.* **2017**, *28*, 1704202.
- (39) Millero, F. J. *Chemical Oceanography* (4th Ed.) CRC Press: Boca Raton, FL, **2013**.
- (40) Lewis E.; Wallace D. CO2SYS-Program developed for the CO₂ system calculations. ORNL/CDIAC-105. Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, US Department of Energy, Oak Ridge, Tennessee, US, **2006**.
- (41) Blue, C. R.; Giuffrè, A.; Mergelsberg, S.; Han, N.; De Yoreo, J. J.; Dove, P. M. Chemical controls on the magnesium content of amorphous calcium carbonate *Geochim. Cosmochim. Acta* **2017**, *196*, 179–196.
- (42) Wang, D.; Hamm, L. M.; Bodnar, R. J.; Dove, P. M. Raman spectroscopic characterization of the magnesium content in amorphous calcium carbonates *J. Raman Spectrosc.* **2012**, *43*, 543–548.
- (43) DeCarlo, T. M.; D’Olivo, J. P.; Foster, T.; Holcomb, M.; Becker, T.; McCulloch, M. T. Coral calcifying fluid aragonite saturation states derived from Raman spectroscopy *Biogeosciences*, **2017**, *14*, 5253–5269.
- (44) Tobler, D. J.; Blanco, J. D. R.; Dideriksen, K.; Sand, K. K.; Bovet, N. The effect of aspartic acid and glycine on amorphous calcium carbonate (ACC) structure , stability and crystallization *Proc. Earth Plan. Science*, **2014**, *10*, 143–148.
- (45) Tlili, M. M.; Amor, M. B.; Gabrielli, C.; Joiret, S.; Maurin, G.; Rousseau, P. Characterization of CaCO₃ hydrates by micro-Raman spectroscopy *J. Raman Spectrosc.* **2001**, *33*, 10–16.
- (46) Politi, B. Y.; Levi-Kalishman, Y.; Raz, S.; Wilt, F.; Addadi, L.; Weiner, S.; Sagi, I. Structural characterization of the transient amorphous calcium carbonate precursor phase in sea urchin embryos *Adv. Funct. Mater.* **2006**, *16*, 1289–1298.
- (47) Herman, R. G.; Bogdan, C. E.; Sommer, A. J.; Simpson, D. R. Discrimination among carbonate minerals by Raman spectroscopy using the laser microprobe *Appl. Spectrosc.* **1987**, *41*, 437–440.
- (48) Radha, A. V.; Fernandez-Martinez, A.; Hu, Y.; Jun, Y.-S.; Waychunas, G. A.; Navrotsky, A. Energetic and structural studies of amorphous Ca_{1-x}Mg_xCO₃·nH₂O (0≤x≤1) *Geochim. Cosmochim. Acta* **2012**, *90*, 83–95.
- (49) Reeder, R. J.; Tang, Y.; Schmidt, M. P.; Kubista, L. M.; Cowan, D. F.; Philips, B. L. Characterization of structure in biogenic amorphous calcium carbonate: Pair distribution function and nuclear magnetic resonance studies of lobster gastrolith *Cryst. Growth Des.* **2013**, *13*, 1905–1914.
- (50) Politi, Y.; Batchelor, D. R.; Zaslansky, P.; Chmelka, B. F.; Weaver, J. C.; Sagi, I.; Weiner, S.; Addadi, L. Role of magnesium ion in the stabilization of biogenic amorphous calcium carbonate: A structure - function investigation *Chem. Mater.* **2010**, *12*, 161–166.
- (51) Purgstaller, B.; Mavromatis, V.; Immenhauser, A.; Dietzel, M. Transformation of Mg-bearing amorphous calcium carbonate to Mg-calcite – In situ monitoring *Geochim. Cosmochim. Acta* **2016**, *174*, 180–195.

- (52) Boskey, A. L.; Posner, A. S. Conversion of amorphous calcium phosphate to microcrystalline hydroxyapatite *J. Phys. Chem.* **1973**, *77*, 2313–2317.
- (53) Rodriguez-Blanco, J. D.; Shaw, S.; Bots, P.; Benning, L. G. The role of pH and Mg on the stability and crystallization of amorphous calcium carbonate *J. Alloys Compd.* **2012**, *536S*, S477–S479.
- (54) Raz, B. S.; Hamilton, P. C.; Wilt, F. H.; Weiner, S.; Addadi, L. The transient phase of amorphous calcium carbonate in sea urchin larval spicules: The involvement of proteins and magnesium ions in its formation and stabilization *Adv. Funct. Mater.* **2003**, *13*, 480–486.
- (55) Kim, Y.-Y.; Carloni, J. D.; Demarchi, B.; Sparks, D.; Reid, D. G.; Kunitake, M. E.; Tang, C. C.; Duer, M. J.; Freeman, C. L.; Pokroy, B.; Penkman, K.; Harding, J. H.; Estroff, L. A.; Baker, S. P.; Meldrum, F. C. Tuning hardness in calcite by incorporation of amino acids *Nat. Mater.* **2016**, *15*, 903–910.
- (56) Von Euw, S.; Zhang, Q.; Manichev, V.; Murali, N.; Gross, J.; Feldman, L. C.; Gustafsson, T.; Flach, C.; Mendelsohn, R.; Falkowski, P. G. Biological control of aragonite formation in stony corals *Science* **2017**, *356*, 933–938.
- (57) Sevilgen, D. S.; Venn, A. A.; Hu, M. Y.; Tambutté, E.; de Beer, D.; Planas-Bielsa, V.; Tambutté, S. Full in vivo characterization of carbonate chemistry at the site of calcification in corals *Sci. Adv.* **2019**, *5*, doi:10.1126/sciadv.aau7447.
- (58) Decarlo, T. M.; Comeau, S.; Cornwall, C. E.; McCulloch, M. T. Coral resistance to ocean acidification linked to increased calcium at the site of calcification *Proc. R. Soc. B Biol. Sci.* **2018**, *285*, 20180564.
- (59) Decarlo, T. M. Characterizing coral skeleton mineralogy with Raman spectroscopy *Nat. Commun.* **2018**, *9*, doi:10.1038/s41467-018-07601-3.
- (60) Akiva, A.; Neder, M.; Kahil, K.; Gavriel, R.; Pinkas, I.; Goobes, G.; Mass, T. Minerals in the pre-settled coral *Stylophora pistillata* crystallize via protein and ion changes *Nat. Commun.* **2018**, *9*, doi:10.1038/s41467-018-04285-7.
- (61) de Nooijer, L. J.; Spero, H. J.; Erez, J.; Bijma, J.; Reichart, G. J. Biomineralization in perforate foraminifera *Earth-Science Rev.* **2014**, *135*, 48–58.

The characteristics and biological relevance of inorganic amorphous calcium carbonate (ACC) precipitated from seawater

David Evans, Paul B. Webb, Kirsty Penkman, Roland Kröger, and Nicola Allison



ACC precipitated from normal seawater is formed at a $[\text{CO}_3^{2-}]$ of 10 ± 2 mM, exceeding that at the calcification site of marine organisms reported to utilise an ACC precursor such as corals. The only investigated method of lowering the necessary [DIC] for precipitation was to reduce the seawater Mg/Ca ratio, raising the possibility that this is an important component of utilising an ACC pathway.